

Introduction: During a previous project PhD Candidate Christopher Kemp created 18 (6 treatments x 2-3 replicates) mesocosms (each of 1,000 cubic liters in size) for the purpose of quantifying the differences in growth of larval walleye at different zooplankton densities in Sandusky Bay, Ohio. Densities of zooplankton in the mesocosms consisted of 6 treatments created by pumping water through a zooplankton net (80 mesh size). The X treatment (Fig. 1) was created by filling mesocosms with unfiltered lake water, this represented the ambient zooplankton density (X) in Lake Erie during the 2019 spawning season. The other 5 treatments were created by filtering a proportion of the time required to fill the X treatment: X*5, X*0.5, X*0.25, X*0.1, X*0 (control; all water filtered). Three trials using the above design were conducted with larval Walleye beginning of three size classes (small = ~ 11 - 13 mm, medium = ~ 15 - 18 mm and large = ~ 21 - 25 mm). Walleye were placed in the mesocosms and allowed to feed on zooplankton for 3 days, at which time they were removed from the mesocosms and preserved in 70% ethanol. Replicate zooplankton samples were taken prior to addition of larval Walleye to the mesocosms and after the larvae were removed. Beginning and final Zooplankton biomass was quantified following the 100 species method (Mack et al., 2012). While Mr. Kemp has quantified larval fish growth using otolith incremental analyses (not explained here), I compared the actual diets and prey consumption at the end of the 3-day experiments.

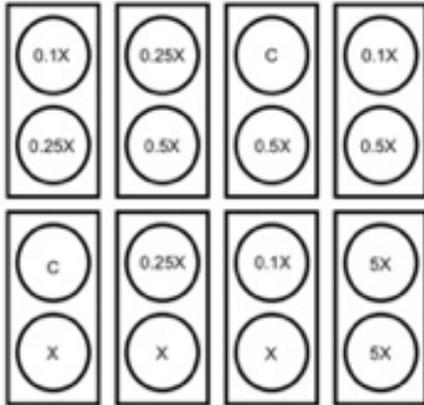


Figure 1. Mesocosm design in Sandusky Bay, Ohio to determine effects of zooplankton density on larval Walleye growth. Each mesocosm held 1m³ of water. ‘0.5X’ indicates that half the water used to fill the mesocosm was filtered to remove 50% of the ambient zooplankton, where ‘X’ indicates ambient zooplankton densities. ‘C’ indicates a control treatment, where all zooplankton were filtered and removed. Note that only two replicate mesocosms were sampled for the control (C) and 5X conditions to optimize statistical design.

Procedure: Preserved larvae harvested at the end of each experiment were measured to total length (TL mm). The entire gut of the larvae was extracted from behind the pectoral fins to the anus under a dissecting microscope. Diet items were quantified under a dissecting microscope and measured (for the purpose of conversion to biomass following Culver et al., (1985)). These data were then analyzed to assess the relationship between available zooplankton biomass (in mesocosms) and biomass consumed.

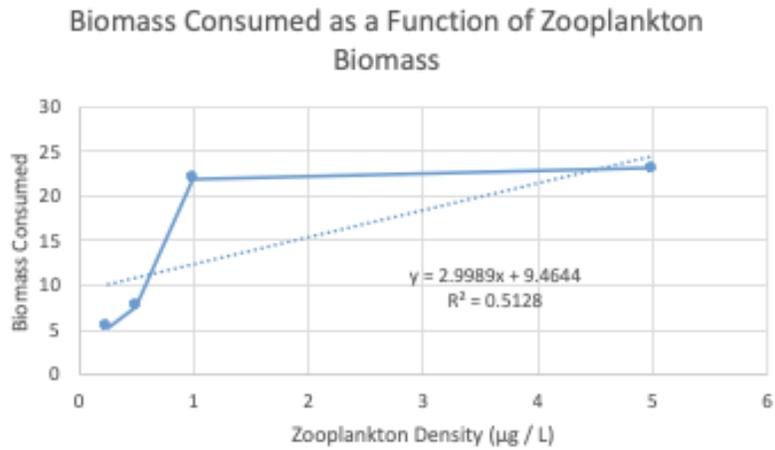


Figure 1. The Functional Response: Biomass consumed as a function of zooplankton biomass in mesocosms

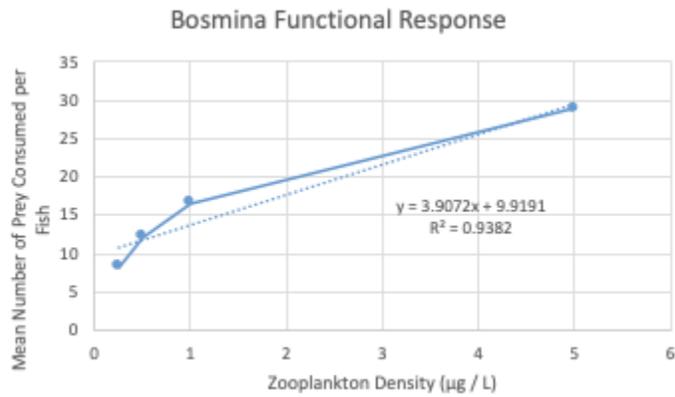


Figure 2. Walleye larvae consumption of Bosmina as a function of zooplankton biomass

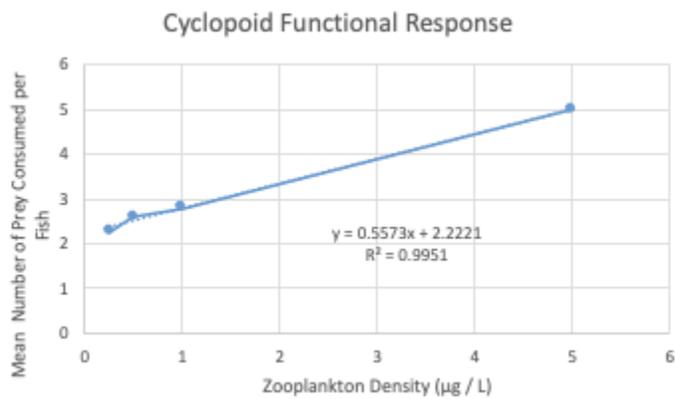


Figure 3. Walleye larvae consumption of cyclopoid copepods as a function of zooplankton biomass

References:

Begon, M., Harper, J. L., & Townsend, C. R. 1986. *Ecology: Individuals, populations and communities*. Blackwell scientific publications.

Chesson, J. 1983. The estimation and analysis of preference and its relationship to foraging models. *Ecology*. 1983;64(5):1297–1304. doi: 10.2307/1937838.

Culver, D. A., Boucherle, M. M., Bean, D. J., & Fletcher, J. W. 1985. Biomass of freshwater crustacean zooplankton from length–weight regressions. *Canadian Journal of Fisheries and Aquatic Sciences*, 42(8), 1380-1390.

- Hoxmeier, R. J. H., D. H. Wahl, M. L. Hooe, and C. L. Pierce. 2004. Growth and Survival of Larval Walleyes in Response to Prey Availability. *Transactions of the American Fisheries Society* 133:45–54.
- Mack, H. R., J. D. Conroy, K. A. Blocksom, R. A. Stein, and S. A. Ludsin. 2012. A comparative analysis of zooplankton field collection and sample enumeration methods. *Limnology and Oceanography: Methods* 10:41–53.
- Madon, S., and D. Culver. 1993. Bioenergetics Model for Larval and Juvenile Walleyes - an in-Situ Approach with Experimental Ponds. *Transactions of the American Fisheries Society* 122:797–813.
- McElman, J. F., and E. K. Balon. 1979. Early ontogeny of walleye, *Stizostedion vitreum*, with steps of saltatory development. *Environmental Biology of Fishes* 4:309–348.
- Mion, J. B., R. A. Stein, and E. A. Marschall. 1998. River Discharge Drives Survival of Larval Walleye. *Ecological Applications* 8:88–103.
- Zorn, T. G. Z., D. B. Hayes, D. E. McCullough, and N. M. Watson. 2020. Crustacean zooplankton available for larval walleyes in a Lake Michigan embayment. *Journal of Great Lakes Research* 1:1491–1499.